

Testimony of Daniel Paul Perez, Co-Founder & Director Emeritus and past Chairman, President & Chief
Executive Officer, Chief Scientific Officer, FSHD Society before
U.S. Senate Appropriations Subcommittee on Labor, HHS, Education and Related Agencies
June 24, 2021

Honorable Chairwoman Murray, Ranking Member Blunt, and distinguished members of the Subcommittee, thank you for the opportunity to testify. We are requesting the FY2022 appropriation of an *amount* of **\$33 million** for the *agency* U.S. DHHS National Institutes of Health (NIH) *program* on research specifically directed at facioscapulohumeral disease and facioscapulohumeral muscular dystrophy (*hereafter called FSHD*).

FSHD is a heritable disease and one of the most common neuromuscular disorders with a prevalence of 1:8,000.¹ It affects 934,000 children and adults of both sexes worldwide. FSHD is characterized by progressive loss of skeletal muscle strength that is asymmetric in pattern and widely variable. Muscle weakness typically starts at the face, shoulder girdle and upper arms, often progressing to the legs, torso and other muscles. In addition to affecting muscle it can bring with it respiratory failure and breathing issues⁶², mild-profound hearing loss, eye problems and cardiac bundle blockage and arrhythmias⁷⁹. FSHD causes significant disability and death according the U.S. Centers for Disease Control and Prevention (CDC), National Center on Birth Defects and Developmental Disabilities, Atlanta, Georgia and others.^{80,81}

FSHD is associated with epigenetic changes on the tip of human chromosome 4q35 in the D4Z4 DNA macrosatellite repeat array region leading to an inappropriate gain of expression (function) of the D4Z4-embedded double homeobox 4 (DUX4) gene². DUX4 is a transcription factor that kick starts the embryonic genome during the 2- to 8-cell stage of development³⁻⁵. Ectopic expression of DUX4 in skeletal muscle is associated with the disease and the disease's pathophysiology that leads to muscle death. DUX4 is never expressed in 'healthy' muscle. FSHD has had few clinical trials⁶⁻¹⁰, and currently there is no cure or therapeutic option available to patients. DUX4 requires and needs to activate its direct transcriptional targets for DUX4-induced gene aberration and muscle toxicity¹¹⁻²⁴. The genetics of FSHD are so remarkable, that NIH Director Dr. Francis Collins said on the front page of the New York Times, "If we were thinking of a collection of the genome's greatest hits, this [FSHD] would go on the list."⁷⁸

Blocking DUX4's DNA, DUX4's RNA or DUX4's protein ability to activate its targets has profound therapeutic relevance²⁵. The FSHD scientific community has in recent years pioneered inroads to treating FSHD using the enormous potential of genomic sequencing, genomic medicine, gene editing and next generation diagnostics. Table 1 lists a dozen approaches detailed in thirty-eight proof-of-concept publications that molecular and genetic treatment approaches work in cellular and animal models for FSHD. All with the central paradigm of the reduction of: DUX4, DUX4 expression, DUX4 protein activity, or the effects of DUX4-mediated toxicity. Strategies include modulating DUX4 repressive pathways, targeting DUX4 mRNA, DUX4 protein, or cellular downstream effects of DUX4 expression. Simply unfathomable as to why NIH funding in this area is not increasing with the pace of discovery.

TABLE 1: Genetic Approaches with Potential to Treat FSHD

- Targeting the DUX4 gene itself by repression using CRISPR/dSaCas9 or CRISPR/dCas9-KRAB;
- Targeting and correcting the FSHD2 SMCHD1 gene mutation with CRISPR/Cas9;
- Knockdown and silencing of the DUX4 gene by going after DUX4 mRNA with antisense oligonucleotides and with RNA interference; U7-asDUX4 snRNAs;
- Targeting DUX4 protein expression using through DNA aptamers; proteins homologous to DUX4; and DNA decoys;
- Going after and controlling expression target downstream [post-expression] of DUX4;
- Going after genetic modifiers of DUX4 expression and DUX4-mediated toxicity between the DUX4 gene and DUX4 mRNA; G-quadruplexes (GQs); and
- Targeting proteins that perturb DUX4-mediated toxicity or secondary features of FSHD pathology.²⁶⁻⁶³

The clinical trials readiness priorities remain similar to last year's testimony. The FSHD scientific community has listed emphasis areas as: 1.) clinical trials readiness infrastructure and therapeutics; 2.) direct and surrogate

biomarkers; 3.) genetic testing, genetics and epigenetics; 4.) imaging and outcome measures; and, 5.) registries and patient focused and reported outcomes.⁷³ The way to measuring disease progression and the effectiveness and safety of drugs remains deep and hard-going for industry, clinical partners and patients.

Serendipitously, new NextGen genomic sequencing and diagnostic technologies, as well as gene-targeted therapeutic approaches have emerged that will be game changing for FSHD patients and families. Understanding one's disease or condition is key for both mental and physical health. This can also aid with family and life planning decisions. With certainty many barriers to matching FSHD disease severity to outcome measures would rapidly fall. We could better align drug and therapeutic modalities with proper phenotypic/genotypic silos of FSHD based on repeat unit, methylation ranges and other requisites for FSHD. The current testing approach in the US, albeit excellent, has created a drag on the momentum towards clinical trials. With therapies on the way, identifying asymptomatic carriers and those that will decades later have later onset or mild symptoms, will allow us to then halt the disease in its early formative stages.^{64,66-69,72}

Recently in 2021, two excellent papers were published on FSHD and DUX4. Both were outstanding – one was using Oxford Nanopore long read sequencing of direct-RNA to locate DUX4 gene targets and the other was a careful study of DUX4 expression in its endogenous [native] form versus the more common recombinant [created] form used in the laboratory.^{70,71} As I read, I asked myself of each: “does this tell us anything more about what DUX4’s function is? No. How DUX4 works? Nada. Or how DUX4 causes FSHD pathophysiology? Nothing at all. How and if DUX4 itself is toxic to skeletal muscle? Zilch. If all research using FSHD transgenic cells in animals is simply result of an artifact? Not sure now.” Both papers yield the same thought: though DUX4 is the prime therapeutic target -- we know next to nothing about it. It is still a complete black box; yet the central focus for FSHD therapy. Questions and areas of research interest emerge from these publications and allied considerations; flowing fast -- each one hypothesis worthy of several NIH grants. “Is DUX4 cytotoxicity pathogenic in vivo? How does expression of DUX4 lead to muscle loss? What is the role of non-muscle cells in FSHD pathology? Can muscle pathology be stopped once it has started (as visualized via MRI images) or is it too late? How is DUX4 bursting regulated in vivo? What other cell types express DUX4 in FSHD and/or healthy individuals? Does the DUX4 mRNA play a nuclear role in FSHD? Are there noncoding RNA roles for DUX4? Are DUX4 induced protein aggregates cause or consequence for FSHD? Does autoimmunity play a role in FSHD? Are there other DUX4-dependent therapeutic targets?” NIH should certainly encourage proposals here. New data/information generated on the basic mechanism of DUX4 and how it causes muscle disease has the potential to focus the design of future clinical trials on muscles and measurements that will increase the rigor of the design and decrease the number of individuals necessary for initial tests of drug activity. It is absolutely necessary to increase our resolution, clarity and understanding of what DUX4 is and what it does to muscle in FSHD. The gains in this area will effectively unpin or untether FSHD from the difficulty category of “slowly progressing neuromuscular diseases remaining recalcitrant” to timely ascertainment that a clinical intervention can work.

Your Subcommittee and **Congress in partnership with NIH, patients and scientists have made truly outstanding progress in understanding and treating the nine major types of muscular dystrophy through the Muscular Dystrophy Community Assistance, Research and Education Amendments of 2001 (MD-CARE Act, Public Law 107-84).** Since passing the MD CARE Act in 2001, NIH funding for FSHD has not kept up pace with scientific opportunities listed herein. The NIH is the principal worldwide source of funding of research on FSHD. Currently active projects are \$16.554 million FY2022 (current actual 23June2021), a 21% portion of the estimated \$80 million spent on all muscular dystrophies. (source: NIH Research Portfolio Online Reporting Tools (RePORT) keyword ‘FSHD or facioscapulohumeral or landouzy-dejerine’).

FSHD Research Dollars (in millions) & FSHD as a Percentage of Total NIH Muscular Dystrophy Funding

Sources: NIH/OD Budget Office & NIH OCPL & NIH RePORT RCDC (e=estimate, a=actual)

Fiscal Year	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021
All MD (\$ millions)	\$83	\$86	\$75	\$75	\$76	\$78	\$77	\$79	\$81	\$81	\$83	\$88e	80e
FSHD (\$ millions)	\$5	\$6	\$6	\$5	\$5	\$7	\$8	\$9	\$11	\$11	\$10	\$11e	\$10e
FSHD (% total MD)	6%	7%	8%	7%	7%	9%	10%	11%	14%	14%	12%	13%	13%

We request for FY2022, a doubling of the NIH FSHD research portfolio to \$33 million. At this moment in time, FSHD needs an infusion of NIH grants both submitted and funded. NIH needs to increase funding by adding exploratory / developmental research grants (parent R21) and research project grants (parent R01) in areas outlined by experts both in

this testimony and in the 2015 DHHS NIH MD Plan.⁷⁷ NIH can issue targeted funding announcements covering FSHD. These efforts will help NIH receive more grant applications. This is NIH's wheelhouse and forte without a doubt.

Madam Chairman, this is my sixty-second testimony before the U.S. Congress' Appropriations Subcommittee on this matter. My FSHD is a strong fort; it has lasted my lifetime of fifty-nine years. That is a long time to live with a disease of this burden.⁸⁰ I hope with your help and action to be able to outlive my disease. I need your help, my friends and fellow FSHD patients and families need your help. Please implore NIH to double funding on FSHD and kindly remember that our lives matter. Madam Chairman, thank you again for your help and efforts.

REFERENCES

1. Deenen, J. C. W. et al. Population-based incidence and prevalence of facioscapulohumeral dystrophy. *Neurology* 83, 1056–9 (2014).
2. Wang, L. H. & Tawil, R. Facioscapulohumeral Dystrophy. *Curr. Neurol. Neurosci. Rep.* 16, 66 (2016).
3. Hendrickson, P. G. et al. Conserved roles of mouse DUX and human DUX4 in activating cleavage-stage genes and MERVL/HERVL retrotransposons. *Nat. Genet.* 49, 925–934 (2017).
4. Whiddon, J. L., Langford, A. T., Wong, C.-J., Zhong, J. W. & Tapscott, S. J. Conservation and innovation in the DUX4-family gene network. *Nat. Genet.* 49, 935–940 (2017).
5. De Iaco, A. et al. DUX-family transcription factors regulate zygotic genome activation in placental mammals. *Nat. Genet.* 49, 941–945 (2017).
6. Tawil, R. et al. A pilot trial of prednisone in facioscapulohumeral muscular dystrophy. FSHDY Group. *Neurology* 48, 46–9 (1997).
7. Passerieux, E. et al. Effects of vitamin C, vitamin E, zinc gluconate, and selenomethionine supplementation on muscle function and oxidative stress biomarkers in patients with facioscapulohumeral dystrophy: a double-blind randomized controlled clinical trial. *Free Radic. Biol. Med.* 81, 158–69 (2015).
8. Kissel, J. T. et al. Randomized, double-blind, placebo-controlled trial of albuterol in facioscapulohumeral dystrophy. *Neurology* 57, 1434–40 (2001).
9. Elsheikh, B. H. et al. Pilot trial of diltiazem in facioscapulohumeral muscular dystrophy. *Neurology* 68, 1428–9 (2007).
10. Wagner, K. R. et al. A phase I/II trial of MYO-029 in adult subjects with muscular dystrophy. *Ann. Neurol.* 63, 561–71 (2008).
11. Rickard, A. M., Petek, L. M. & Miller, D. G. Endogenous DUX4 expression in FSHD myotubes is sufficient to cause cell death and disrupts RNA splicing and cell migration pathways. *Hum. Mol. Genet.* 24, 5901–14 (2015).
12. Sandri, M. et al. Caspase 3 expression correlates with skeletal muscle apoptosis in Duchenne and facioscapulo human muscular dystrophy. A potential target for pharmacological treatment? *J. Neuropathol. Exp. Neurol.* 60, 302–12 (2001).
13. Block, G. J. et al. Wnt/β-catenin signaling suppresses DUX4 expression and prevents apoptosis of FSHD muscle cells. *Hum. Mol. Genet.* 22, 4661–72 (2013).
14. Statland, J. M. et al. Immunohistochemical Characterization of Facioscapulohumeral Muscular Dystrophy Muscle Biopsies. *J. Neuromuscul. Dis.* 2, 291–299 (2015).
15. Rickard, A. M., Petek, L. M. & Miller, D. G. Endogenous DUX4 expression in FSHD myotubes is sufficient to cause cell death and disrupts RNA splicing and cell migration pathways. *Hum. Mol. Genet.* 24, 5901–14 (2015).
16. Kowaljow, V. et al. The DUX4 gene at the FSHD1A locus encodes a pro-apoptotic protein. *Neuromuscul. Disord.* 17, 611–23 (2007).
17. Bosnakovski, D. et al. An isogenetic myoblast expression screen identifies DUX4-mediated FSHD-associated molecular pathologies. *EMBO J.* 27, 2766–79 (2008).
18. Wallace, L. M. et al. DUX4, a candidate gene for facioscapulohumeral muscular dystrophy, causes p53-dependent myopathy in vivo. *Ann. Neurol.* 69, 540–52 (2011).
19. Geng, L. N. et al. DUX4 activates germline genes, retroelements, and immune mediators: implications for facioscapulohumeral dystrophy. *Dev. Cell* 22, 38–51 (2012).
20. Yao, Z. et al. DUX4-induced gene expression is the major molecular signature in FSHD skeletal muscle. *Hum. Mol. Genet.* 23, 5342–52 (2014).
21. Homma, S., Beermann, M., Lou, Boyce, F. M. & Miller, J. B. Expression of FSHD-related DUX4-FL alters proteostasis and induces TDP-43 aggregation. *Ann. Clin. Transl. Neurol.* 2, 151–66 (2015).
22. Jagannathan, S. et al. Model systems of DUX4 expression recapitulate the transcriptional profile of FSHD cells. *Hum. Mol. Genet.* 25, 4419–4431 (2016).
23. Jones, T. I. et al. Facioscapulohumeral muscular dystrophy family studies of DUX4 expression: evidence for disease modifiers and a quantitative model of pathogenesis. *Hum. Mol. Genet.* 21, 4419–30 (2012).
24. Campbell AE, Shadle SC, Jagannathan S, Lim JW, Resnick R, Tawil R, van der Maarel SM, Tapscott SJ. NuRD and CAF-1-mediated silencing of the D4Z4 array is modulated by DUX4-induced MBD3L proteins. *Elife.* 2018 Mar 13;7. pii: e31023. doi: 10.7554/eLife.31023 (2018).
25. Jagannathan S1,2,3, Ogata Y4, Gaffen PR4, Tapscott SJ3, Bradley RK1. Quantitative proteomics reveals key roles for post-transcriptional gene regulation in the molecular pathology of facioscapulohumeral muscular dystrophy. *Elife.* 2019 Jan 15;8. pii: e41740. doi: 10.7554/eLife.41740 (2019).
26. Vanderplanck, C., Anseau, E., Charron, S., Stricwant, N., Tassin, A., Laoudi-Chenivesse, D., et al. The FSHD atrophic myotube phenotype is caused by DUX4 expression. *PLoS One* 6, e26820. doi:10.1371/journal.pone.0026820 (2011).
27. Wallace, L. M., Liu, J., Domire, J. S., Garwick-Coppens, S. E., Guckes, S. M., Mendell, J. R., et al. RNA interference inhibits DUX4-induced muscle toxicity in vivo: implications for a targeted FSHD therapy. *Mol. Ther.* 20, 1417–1423. doi:10.1038/mt.2012.68 (2012)
28. Pandey, S. N., Lee, Y. C., Yokota, T., and Chen, Y. W. Morpholino treatment improves muscle function and pathology of Pitx1 transgenic mice. *Mol. Ther.* 22, 390–396. doi:10.1038/mt.2013.263 (2014).
29. Lim, J. W., Snider, L., Yao, Z., Tawil, R., Van Der Maarel, S. M., Rigo, F., et al. DICER/AGO-dependent epigenetic silencing of D4Z4 repeats enhanced by exogenous siRNA suggests mechanisms and therapies for FSHD. *Hum. Mol. Genet.* 24, 4817–4828. doi:10.1093/hmg/ddv206 (2015).
30. Marsollier, A. C., Ciszewski, L., Mariot, V., Popplewell, L., Voit, T., Dickson, G., et al. Antisense targeting of 3' end elements involved in DUX4 mRNA processing is an efficient therapeutic strategy for facioscapulohumeral dystrophy: a new gene-silencing approach. *Hum. Mol. Genet.* 25, 1468–1478. doi:10.1093/hmg/ddw015 (2016).
31. Himeda CL, Jones, et al. CRISPR/dCas9-mediated Transcriptional Inhibition Ameliorates the Epigenetic Dysregulation at D4Z4 and Represses DUX4-fl in FSH Muscular Dystrophy. *Mol Ther.* Mar;24(3):527-35. epub 2015 Nov 3. (2016).
32. Chen JC, King OD, Zhang Y, et al. Morpholino-mediated Knockdown of DUX4 Toward Facioscapulohumeral Muscular Dystrophy Therapeutics. *Molecular Therapy.* 24 (8):1405-1411. doi:10.1038/mt.2016.1118. (2016).
33. Anseau, E., Vanderplanck, C., Wauters, A., Harper, S. Q., Coppée, F., and Belayew, A. Antisense oligonucleotides used to target the DUX4 mRNA as therapeutic approaches in FacioscapuloHumeral muscular dystrophy (FSHD). *Genes* 8, 93. doi:10.3390/genes8030093 (2017).
34. Bosnakovski, D., Toso, E. A., Hartweck, L. M., Magli, A., Lee, H. A., Thompson, E.R., et al. The DUX4 homeodomains mediate inhibition of myogenesis and are functionally exchangeable with the Pax7 homeodomain. *J. Cell Sci.* 130, 3685–3697. doi:10.1242/jcs.205427 (2017).
35. Marsollier AC, Joubert R, Mariot V, Dumonceaux J. Targeting the Polyadenylation Signal of Pre-mRNA: A New Gene Silencing Approach for Facioscapulohumeral Dystrophy. *Int J Mol Sci.* May 3;19(5). pii: E1347. doi: 10.3390/ijms19051347. Review. (2018).
36. Lee JK, Bosnakovski D, Toso EA, Dinh T, Banerjee S, Bohl TE, Shi K, Orellana K, Kyba M, Aihara H. Crystal Structure of the Double Homeodomain of DUX4 in Complex with DNA. *Cell Rep.* Dec 11;25(11):2955-2962.e3. doi: 10.1016/j.celrep.2018.11.060. (2018).
37. Lim, K. R. Q., and Yokota, T. Invention and early history of exon skipping and splice modulation, in Exon skipping and inclusion therapies: methods and protocols. Editors T. Yokota and R. Maruyama (New York, NY: Springer), 3–30. (2018).
38. Himeda CL, Jones TI, Virbasius CM, Zhu LJ, Green MR, Jones PL. Identification of Epigenetic Regulators of DUX4-fl for Targeted Therapy of Facioscapulohumeral Muscular Dystrophy. *Mol Ther.* Jul 5;26(7):1797-1807. doi: 10.1016/j.yimthe.2018.04.019. Epub 2018 Apr 26. (2018).
39. Wallace, L. M., Saad, N. Y., Pyne, N. K., Fowler, A. M., Eidahl, J. O., Domire, J. S., et al. Pre-clinical safety and off-target studies to support translation of AAV-mediated RNAi therapy for FSHD. *Mol. Ther. Methods Clin. Dev.* 8, 121–130. doi:10.1016/j.omtm.2017.12.005 (2018).
40. Giesing CR, Wallace LM, Heller KN, Eidahl JO, Saad NY, Fowler AM, Pyne NK, Al-Kharsan M, Rashnonejad A, Chermahini GA, Domire JS, Mukweyi D, Garwick-Coppens SE, Guckes SM, McLaughlin KJ, Meyer K, Rodino-Klapac LR, Harper SQ. AAV-mediated follistatin gene therapy improves functional outcomes in the TIC-DUX4 mouse model of FSHD. *JCI Insight.* 2018 Nov 15;3(22). pii: 123538. doi: 10.1172/jci.insight.123538. (2018).
41. Jones, T., and Jones, P. L. (2018). A cre-inducible DUX4 transgenic mouse model for investigating facioscapulohumeral muscular dystrophy. *PLoS One* 13, e0192657. doi:10.1371/journal.pone.0192657 (2018).
42. Mitsuhashi, H., Ishimaru, S., Homma, S., Yu, B., Honma, Y., Beermann, M. L., et al. Functional domains of the FSHD-associated DUX4 protein. *Biol. Open* 7, bio033977. doi:10.1242/bio.033977 (2018).

43. Goossens, R., van den Boogaard, M. L., Lemmers, R. J. L. F., Balog, J., van der Vliet, P. J., Willemsen, I. M., et al. Intronic SMCHD1 variants in FSHD: testing the potential for CRISPR-Cas9 genome editing. *J. Med. Genet.* 56, 828–837. doi:10.1136/jmedgenet-2019-106402 (2019).
44. Dion C, Roche S, Laberthonniere C, Broucqsaunt N, Mariot V, Xue S, Gurzau AD, Nowak A, Gordon CT, Gaillard MC, El-Yazidi C, Thomas M, Schlupp-Robaglia A, Missirian C, Malan V, Rabi L, Sefiani A, Wollnik B, Binetruy B, Salort Campana E, Attarian S, Bernard R, Nguyen K, Amiel J, Dumonceaux J, Murphy JM, Déjardin J, Blewitt ME, Reversade B, Robin JD, Magdinier F. SMCHD1 is involved in de novo methylation of the DUX4-encoding D4Z4 macrosatellite. *Nucleic Acids Res.* 2019 Jan 30. doi: 10.1093/nar/gkz005. [Epub ahead of print] (2019).
45. Jagannathan S, Ogata Y, Gafken PR, Tapscott SJ, Bradley RK. Quantitative proteomics reveals key roles for post-transcriptional gene regulation in the molecular pathology of facioscapulohumeral muscular dystrophy. *Elife.* 2019 Jan 15;8:e41740. doi: 10.7554/eLife.41740. PMID: 30644821 (2019).
46. Lim, K. R. Q., and Yokota, T. (2020). Invention and early history of gapmers. *Methods Mol. Biol.* 2176, 3–19. doi:10.1007/978-1-0716-0771-8_1 (2020).
47. Ciszewski L, Lu-Nguyen N, Slater A, Brennan A, Williams HEL, Dickson G, Searle MS, Popplewell L. G-quadruplex ligands mediate downregulation of DUX4 expression. *Nucleic Acids Res.* 2020 May 7;48(8):4179–4194. doi: 10.1093/nar/gkaa146. (2020).
48. Lim KRQ, Maruyama R, Echigoya Y, Nguyen Q, Zhang A, Khawaja H, Sen Chandra S, Jones T, Jones P, Chen YW, Yokota T. Inhibition of DUX4 expression with antisense LNA gapmers as a therapy for facioscapulohumeral muscular dystrophy. *Proc Natl Acad Sci U S A.* 2020 Jul 14;117(28):16509–16515. doi: 10.1073/pnas.1909649117. (2020).
49. Lim, K. R. Q., Nguyen, Q., and Yokota, T. DUX4 signaling in the pathogenesis of facioscapulohumeral muscular dystrophy. *Int. J. Mol. Sci.* 21, 729. doi:10.3390/ijms21030729
50. Derenne, A., Tassin, A., Nguyen, T. H., De Roeck, E., Jenart, V., Anseau, E., et al. (2020). Induction of a local muscular dystrophy using electroporation in vivo: an easy tool for screening therapeutics. *Sci. Rep.* 10, 11301. doi:10.1038/s41598-020-68135-7 (2020).
51. Bittel, A. J., Sreetama, S. C., Bittel, D. C., Horn, A., Novak, J. S., Yokota, T., et al. Membrane repair deficit in facioscapulohumeral muscular dystrophy. *Int. J. Mol. Sci.* 21, 5575. doi:10.3390/ijms21155575 (2020).
52. Jones TI, Chew GL, Barraza-Flores P, Schreier S, Ramirez M, Wuebbles RD, Burkin DJ, Bradley RK, Jones PL. Transgenic mice expressing tunable levels of DUX4 develop characteristic facioscapulohumeral muscular dystrophy-like pathophysiology ranging in severity. *Skelet Muscle.* Apr 11;10(1):8. doi: 10.1186/s13395-020-00227-4 (2020).
53. Mariot, V., Joubert, R., Marsollier, A.-C., Houdé, C., Voit, T., and Dumonceaux, J. A deoxyribonucleic acid decoy trapping DUX4 for the treatment of facioscapulohumeral muscular dystrophy. *Mol. Ther. Nucleic Acids.* doi:10.1016/j.omtn.2020.10.028 (2020).
54. Cohen, J., DeSimone, A., Lek, M., and Lek, A. Therapeutic approaches in facioscapulohumeral muscular dystrophy. *Trends Mol. Med.* 27, 123–37. doi:10.1016/j.molmed.2020.09.008 (2020).
55. Lek, A., Zhang, Y., Woodman, K. G., Huang, S., DeSimone, A. M., Cohen, J., et al. Applying genome-wide CRISPR-Cas9 screens for therapeutic discovery in facioscapulohumeral muscular dystrophy. *Sci. Transl. Med.* 12, eaay0271. doi:10.1126/scitranslmed.aay0271 (2020).
56. Lemmers RJLF, van der Vliet PJ, Blatnik A, Balog J, Zidar J, Henderson D, Goslinsk R, Tapscott SJ, Voermans NC, Tawil R, Padberg GWAM, van Engelen BG, van der Maarel SM. Chromosome 10q-linked FSHD identifies DUX4 as principal disease gene. *J Med Genet.* 2021 Jan 12; jmedgenet-2020-107041. doi: 10.1136/jmedgenet-2020-107041 (2021).
57. Rashnonejad A, Amini-Chermahini G, Taylor NK, Wein N, Harper SQ. Designed U7 snRNAs inhibit DUX4 expression and improve FSHD-associated outcomes in DUX4 overexpressing cells and FSHD patient myotubes. *Mol Ther Nucleic Acids.* 2020 Dec 10;23:476–486. doi: 10.1016/j.omtn.2020.12.004. eCollection 2021 Mar 5 (2021).
58. Himeida CL, Jones TI, Jones PL. Targeted epigenetic repression by CRISPR/dSaCas9 suppresses pathogenic DUX4-fl expression in FSHD. *Mol Ther Methods Clin Dev.* 2020 Dec 10;20:298–311. doi: 10.1016/j.omtn.2020.12.001. eCollection 2021 Mar 12 (2021).
59. Schätzl T, Kaiser L, Deigner HP. Facioscapulohumeral muscular dystrophy: genetics, gene activation and downstream signaling with regard to recent therapeutic approaches: an update. *Orphanet J Rare Dis.* 2021 Mar 12;16(1):129. doi: 10.1186/s13023-021-01760-1. Review (2021).
60. Lim KRQ, Yokota T. Genetic Approaches for the Treatment of Facioscapulohumeral Muscular Dystrophy. *Front Pharmacol.* 2021 Mar 12;12:642858. doi: 10.3389/fphar.2021.642858. eCollection 2021. Review (2021).
61. Mellion ML, Ronco L, Berends CL, Pagan L, Brooks S, van Esdonk MJ, van Brummelen EMJ, Oduyungbo A, Thompson LA, Hage M, Badrising UA, Raines S, Tracewell WG, van Engelen B, Cadavid D, Groeneveld GJ. Phase 1 clinical trial of losmapimod in facioscapulohumeral dystrophy: Safety, tolerability, pharmacokinetics, and target engagement. *Br J Clin Pharmacol.* 2021 Apr 30. doi: 10.1111/bcp.14884. (2021).
62. Lu-Nguyen N, Malerba A, Herath S, Dickson G, Popplewell L. Systemic antisense therapeutics inhibiting DUX4 expression ameliorates FSHD-like pathology in an FSHD mouse model. *Hum Mol Genet.* 2021 May 13; ddab136. doi: 10.1093/hmg/ddab136. (2021).
63. Das S, Chadwick BP. CRISPR mediated targeting of DUX4 distal regulatory element represses DUX4 target genes dysregulated in Facioscapulohumeral muscular dystrophy. *Sci Rep.* 2021 Jun 15;11(1):12598. doi: 10.1038/s41598-021-92096-0. (2021).
64. Goslinsk RJM, Mul K, van Kernebeek CR, Lemmers RJLF, van der Maarel SM, Schreuder THA, Erasmus CE, Padberg GW, Statland JM, Voermans NC, van Engelen BGM. Early onset as a marker for disease severity in facioscapulohumeral muscular dystrophy. *Neurology.* 2019 Jan 22;92(4):e378–e385. doi: 10.1212/WNL.00000000000006819. Epub 2018 Dec 19 (2019).
65. Henke C, Spiesshoefer J, Kabitz HJ, Herkenrath S, Randerath W, Brix T, Görlich D, Young P, Boentert M. Respiratory muscle weakness in facioscapulohumeral muscular dystrophy. *Muscle Nerve.* 2019 Dec;60(6):679–686. doi: 10.1002/mus.26717. Epub 2019 Oct 23 (2019).
66. Sacconi S, Briand-Suleau A, Gros M, Baudoin C, Lemmers RJLF, Rondeau S, Lagha N, Nigumann P, Cambieri C, Puma A, Chapon F, Stojkovic T, Vial C, Bouhour F, Cao M, Pegoraro E, Petiot P, Behin A, Marc B, Eymard B, Echaniz-Laguna A, Laforet P, Salvati L, Jeanpierre M, Cristofari G, van der Maarel SM. FSHD1 and FSHD2 form a disease continuum. *Neurology.* 2019 May 7;92(19):e2273–e2285. doi: 10.1212/WNL.0000000000007456. Epub 2019 Apr 12 (2019).
67. Salort-Campana E, Fatchi F, Beloribi-Djefafilia S, Roche S, Nguyen K, Bernard R, Cintas P, Solé G, Bouhour F, Ollagnon E, Sacconi S, Echaniz-Laguna A, Kuntzer T, Levy N, Magdinier F, Attarian S. Type 1 FSHD with 6–10 Repeated Units: Factors Underlying Severity in Index Cases and Disease Penetrance in Their Relatives Attention. *Int J Mol Sci.* 2020 Mar 23;21(6):2221. doi: 10.3390/ijms21062221 (2020).
68. Wong CJ, Wang LH, Friedman SD, Shaw D, Campbell AE, Budech CB, Lewis LM, Lemmers RJLF, Statland JM, van der Maarel SM, Tawil RN, Tapscott SJ. Longitudinal measures of RNA expression and disease activity in FSHD muscle biopsies. *Hum Mol Genet.* 2020 Apr 15;29(6):1030–1043. doi: 10.1093/hmg/ddaa031. PMID: 32083293 (2020).
69. Rieken A, Bossler AD, Mathews KD, Moore SA. CLIA Laboratory Testing for Facioscapulohumeral Dystrophy: A Retrospective Analysis. *Neurology.* 2021 Feb 16;96(7):e1054–e1062. doi: 10.1212/WNL.0000000000011412. Epub 2020 Dec 21 (2021).
70. Chau J, Kong X, Viet Nguyen N, Williams K, Ball M, Tawil R, Kiyono T, Mortazavi A, Yokomori K. Relationship of DUX4 and target gene expression in FSHD myocytes. *Hum Mutat.* 2021 Jan 27. doi: 10.1002/humu.24171 (2021).
71. Mitsuhashi S, Nakagawa S, Sasaki-Honda M, Sakurai H, Frith MC, Mitsuhashi H. Nanopore direct RNA sequencing detects DUX4-activated repeats and isoforms in human muscle cells. *Hum Mol Genet.* 2021 Mar 9; ddab063. doi: 10.1093/hmg/ddab063 (2021).
72. Goslinsk RJM, Schreuder THA, Mul K, Voermans NC, Erasmus CE, van Engelen BGM, van Alfen N. Muscle ultrasound is a responsive biomarker in facioscapulohumeral dystrophy. *Neurology.* 2020 Apr 7;94(14):e1488–e1494. doi: 10.1212/WNL.00000000000009211. (2020).
73. Wang LH, Shaw DWW, Faino A, Budech CB, Lewis LM, Statland J, Eichinger K, Tapscott SJ, Tawil RN, Friedman SD. Longitudinal study of MRI and functional outcome measures in facioscapulohumeral muscular dystrophy. *BMC Musculoskelet Disord.* 2021 Mar 10;22(1):262. doi: 10.1186/s12891-021-04134-7 (2021).
74. Greco A, Straasheijm KR, Mul K, van den Heuvel A, van der Maarel SM, Joosten LAB, van Engelen BGM, Puijn GJM. Profiling Serum Antibodies Against Muscle Antigens in Facioscapulohumeral Muscular Dystrophy Finds No Disease-Specific Autoantibodies. *J Neuromuscul Dis.* 2021 May 15. doi: 10.3233/JND-210653. (2021).
75. Karpukhina A, Galkin I, Ma Y, Dib C, Zinovkin R, Pletjushkina O, Chernyakh B, Popova E, Vassetzky Y. Analysis of genes regulated by DUX4 via oxidative stress reveals potential therapeutic targets for treatment of facioscapulohumeral dystrophy. *Redox Biol.* 2021 Jul;43:102008. doi: 10.1016/j.redox.2021.102008. (2021).
76. Banerji CRS, Zammit PS. Pathomechanisms and biomarkers in facioscapulohumeral muscular dystrophy: roles of DUX4 and PAX7. *EMBO Mol Med.* 2021 Jun 21;e13695. doi: 10.15252/emmm.202013695. (2021).
77. Rieff HI, Katz SI et al. The Muscular Dystrophy Coordinating Committee Action Plan for the Muscular Dystrophies. *Muscle Nerve.* 2016 Mar 21. [Epub ahead of print] (2016).
78. Kolata, G., Reanimated 'Junk' DNA Is Found to Cause Disease. *New York Times, Science.* Published online: August 19, 2010 <http://www.nytimes.com/2010/08/20/science/20gene.html>
79. Ducharme-Smith A, Nicolau S, Chahal CAA, Ducharme-Smith K, Rehman S, Jaliparthi K, Khan N, Scott CG, St Louis EK, Liewluck T, Somers VK, Lin G, Brady PA, Milone M. Cardiac Involvement in Facioscapulohumeral Muscular Dystrophy (FSHD). *Front Neurol.* 2021 May 24;12:668180. doi: 10.3389/fneur.2021.668180. (2021).
80. Blokhuis AM, Deenen JCW, Voermans NC, van Engelen BGM, Kievit W, Groothuis JT. The socioeconomic burden of facioscapulohumeral muscular dystrophy. *J Neurol.* 2021 May 27. doi: 10.1007/s00415-021-10591-w. (2021).
81. Wallace B, Smith KT, Thomas S, Conway KM, Westfield C, Andrews JG, Weinert RO, Do TQN, Street N; Muscular Dystrophy Surveillance, Tracking, and Research Network (MD STARnet). Characterization of individuals with selected muscular dystrophies from the expanded pilot of the Muscular Dystrophy Surveillance, Tracking and Research Network (MD STARnet) in the United States. *Birth Defects Res.* 2021 Apr 15;113(7):560–569. doi: 10.1002/bdr2.1764. (2020).