

Testimony of Daniel Paul Perez, Co-Founder & Director Emeritus, FSHD Society before  
U.S. Senate Appropriations Subcommittee on Labor, HHS, Education and Related Agencies  
May 18, 2023

Honorable Chairman Baldwin, Ranking Member Moore Capito, and distinguished members of the Subcommittee, thank you. We are requesting a FY2024 appropriation of **\$32.5 million** to the *agency* U.S. DHHS National Institutes of Health (**NIH**) for facioscapulohumeral muscular dystrophy (*hereafter called* **FSHD**) research *programs*.

I am co-founder, Director emeritus, past -Chairman, -President & CEO, and -CSO of the FSHD Society. As a patient with FSHD, my life's work on FSHD disease and its funding spans nearly every research lab, biotechnology and pharma company working today on FSHD globally. My efforts through the FSHD Society have led to understanding how FSHD1<sup>1</sup> and FSHD2<sup>2,3</sup> work. I was a key architect of the Muscular Dystrophy Community Assistance, Research and Education Amendments of 2001 (MD-CARE Act, Public Law 107-84). I have served on the Muscular Dystrophy Coordinating Committee (MDCC) since its inception and am the longest serving member. MDCC is a Federal Advisory Committee designed to coordinate activities relating to the various forms of muscular dystrophy across the NIH and with other Federal health agencies.

FSHD is a heritable disease and one of the most common neuromuscular disorders with a prevalence of 1:8,000.<sup>4</sup> It affects nearly one million children and adults of both sexes worldwide. FSHD is characterized by progressive loss of skeletal muscle strength that is asymmetric in pattern and widely variable. Muscle weakness typically starts at the face, shoulder girdle and upper arms, often progressing to the legs, torso and other muscles. In addition to affecting any skeletal muscle, it can bring with it respiratory failure and breathing issues<sup>5-7</sup>, mild-profound hearing loss<sup>8</sup>, eye problems and cardiac bundle blockage and arrhythmias<sup>9,10</sup>. FSHD causes significant disability and death according the U.S. Centers for Disease Control and Prevention (CDC), National Center on Birth Defects and Developmental Disabilities, Atlanta, and others<sup>11,12</sup>.

FSHD is associated with epigenetic changes on the tip of human chromosome 4q35, in the D4Z4 DNA macrosatellite repeat array region, leading to an inappropriate gain of expression (function) of the D4Z4-embedded **double homeobox 4 (DUX4) gene**<sup>13-16</sup>. DUX4 is a transcription factor that kick starts the embryonic genome during the 2- to 8-cell stage of development<sup>17-19</sup>. Ectopic expression of DUX4 in skeletal muscle is associated with the disease and the disease's pathophysiology that leads to muscle death. DUX4 is never expressed in 'healthy' muscle. FSHD has had few clinical trials<sup>20-26,27,28</sup>, there is no cure or therapeutic options. DUX4 requires and needs to activate direct transcriptional targets for DUX4-induced gene aberration and muscle toxicity<sup>29-41</sup>.

Our patient community through its fundraising efforts and intensive research-integration has pioneered inroads to treating FSHD using genomic sequencing, genomic medicine, gene editing and next generation diagnostics. All with the goal of reducing DUX4 in its DNA or RNA or protein state, or the effects of DUX4-driven toxicity e.g. modulating DUX4 repressive pathways, targeting DUX4 mRNA, DUX4 protein, or cellular downstream effects of DUX4 expression<sup>44-47</sup>.

Currently active projects listed in NIH RePORT as being applicable to FSHD are \$16.961 million FY2023 (18May2023) vs. \$17.507 million year-ago FY2022 (source: NIH Research Portfolio Online Reporting Tools (RePORT) 'FSHD or facioscapulohumeral or landouzy-dejerine'). The NIH is

currently the principal worldwide source of funding of basic biomedical research on FSHD. Currently annual funding directly dedicated for FSHD listed in NIH RCDC is \$10 million.

**Table 1. FSHD Research Dollars (millions) & FSHD Percentage of Total NIH Muscular Dystrophy Funding**

Sources: NIH/OD Budget Office & NIH OCPL & NIH RePORT / RCDC (e=estimate, a=actual vers=March 31, 2023)

Fiscal Year	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023e	2024e
All MD (\$ millions)	\$75	\$76	\$78	\$77	\$79	\$81	\$81	\$83	\$95	\$82	\$78	\$81	83
FSHD (\$ millions)	\$5	\$5	\$7	\$8	\$9	\$11	\$11	\$10	\$9	\$9	\$10	\$10	10
FSHD (% total MD)	7%	7%	9%	10%	11%	14%	14%	12%	9%	11%	13%	12%	12%

Honorable Chairman, thanks to Congress' work in enacting the MD CARE Act, funding the NIH, and with communications with NIH-leadership and program/legislative staff -- several dystrophies now have FDA approved treatments. NIH has been especially thoughtful with its expenditure and approach to research on muscular dystrophy. In the past year, nearly one hundred peer reviewed publications on FSHD came online. Excellent gains made in expanding basic research, better understanding of natural history and disease heterogeneity, biomarkers and outcomes assessments, therapeutic compounds/interventions and clinical trial design<sup>48-61</sup>. Recent publications further underscore the need for broad and combinatorial therapeutic strategies as it is possible that any single approach may not be successful or not be enough e.g. immunomodulation plus DUX4 inhibition, DUX4 inhibition plus muscle regeneration, FSHD as a myodevelopmental disease ["satellite cell-opathy" a primary satellite cell disease] caused by specific stem cell epigenetically weakly repressed at the D4Z4 repeat needing early intervention<sup>62-67</sup>. The prospect of treating and curing FSHD in the next 12 to 24 months is very real and tangible. More than \$1 billion from investment banks, venture capital, private/philanthropic investors across more than 25 companies recently flowed into companies working on FSHD therapies. Though target DUX4 completely black and white – all are still flying blind when it comes to measuring ectopic DUX4 in human muscle. What emerges -- is, on one end of the spectrum an overarching need for far more basic and mechanistic research e.g. better understanding of the fundamental mechanisms of FSHD disease, including muscle disease. Secondly, on the back-end with getting therapies to patients, is the need to eliminate projects that will never succeed in practice/clinic i.e. 'does this have a pathway to clinic?' Associated with our request for 32.5 million dollars in FY2024, we highlight top priorities, research questions in need of better answers and open-ended areas of investigation in tables 2 and 3 where NIH can improve on its FSHD research spending.

**Table 2. FSHD1 and FSHD2 Broad NIH Cross-Cutting Thematic Areas**

- **Better biomarkers:** we still need to improve on our non-invasive and invasive biomarkers for trial readiness;
- **Clinical variability and disease pattern:** our understanding of the causes for clinical variability in both FSHD1 and FSHD2 is too limited, let alone that we can predict progression;
- Studying the **normal function of DUX4** and how it causes muscle disease can focus design of future clinical trials;
- Investigating if muscle pathology can be reversed once DUX4 expression starts the **pathogenic cascade** in a particular muscle?;
- Exploring if there is a systemic effect of local DUX4 expression that leads to amplification of muscle decline, either **immune or some extracellular signaling**. Can we intercept and control the disease process?;
- Designing **new technologies and methods to track** biological, chemical, mechanical and electrical aspects of muscle function for improving or declining health of FSHD permissive genotype;
- Improving **registries:** need further international integration and sustainability;
- **Prioritizing pathways:** DUX4 activates many different pathways, we don't know if there is any hierarchy in contribution to disease, or different roles;
- **Genetic heterogeneity:** not all patients are explained, population differences, etc.;
- **Molecular/tissue heterogeneity:** do we sufficiently understand the muscle involvement? Other tissues/cell types?;
- **DUX4 dynamics:** what activates DUX4 as all nuclei are (epi)genetically sensitized but only few express DUX4. Is there active silencing (can we take advantage of the 8-cell stage mechanism of DUX4 silencing)? How are the DUX4 signals propagated within fibers, between fibers?;

- **Therapy:** many different routes are being explored e.g. small molecule drugs, biologics, gene transfer/editing, cell/molecular therapies (including oligonucleotides), devices, nutritional and exercise, behavioral and combinatorial therapy. How do we handle this effort with the limited patient population that will be available for trials?;
- Improved **preclinical and large animal models** that accurately reflect mechanisms of disease onset and progression;
- **Genetic and environmental mechanisms** that determine timing of onset / rate of progression in genetically inherited disease;
- **Role of the adaptive and innate immune system** in disease onset and progression;
- **Uniform, reliable and scalable DNA diagnostics**, pre-implantation, prenatal and postnatal;
- Promoting research that helps to increase safety and efficacy of FSHD clinical trials, **addressing toxicity, immune response and efficiency of uptake.**

### Table 3. FSHD1 and FSHD2 Inspirational Research Directions

- Individuals with FSHD1 and FSHD2 are born with DUX4 turned on from day one and it may be years/decades on non-symptomatic disease infiltrating the muscle – accelerate research efforts and ways of screening genetically and identifying and **tracking non-symptomatic disease progression and infiltration;**
- Developing **ways to assess, stop and reverse non-symptomatic disease progression** and infiltration in FSHD muscle e.g. outcome measures herein are strictly biological not clinical functional;
- Help accelerate movement of testable compounds toward clinical trials in FSHD, by the **discovery and generation of tractable markers of both symptomatic and non-symptomatic disease** to provide measurable outcomes;
- Understanding precisely **how FSHD skeletal muscle epigenome protects from FSHD or capitulates** to having active FSHD progression;
- Provide a platform for stakeholders [members of the FSHD research community, pharmaceutical and biotechnology companies interested in developing therapeutics for FSHD, and leading scientists who have successfully exploited model systems in understanding human disease writ large] to **identify viable biomarkers that could be exploited** in clinical trials;
- Identify and elucidate or **engineer specific biomarkers and models** associated with the onset, and progression of FSHD1 and FSHD2;

It is imperative to increase our efforts in the areas mentioned herein -- gains in these areas will help ascertain if therapies are effective, safe and not cost-prohibitive. We request for FY2024, increasing NIH FSHD research funding/appropriation of the standard portfolio to \$32.5 million.

Honorable Chairman, thank you for this opportunity to update you with this testimony.

### REFERENCES

1. Gould T, Jones TI, Jones PL. Precise Epigenetic Analysis Using Targeted Bisulfite Genomic Sequencing Distinguishes FSHD1, FSHD2, and Healthy Subjects. *Diagnostics (Basel)*. 2021 Aug 13;11(8):1469. doi: 10.3390/diagnostics11081469. PMID: 34441403; PMCID: PMC8393475. (2021).
2. Goossens R, Tihaya MS, van den Heuvel A, Tabot-Ndip K, Willemsen IM, Tapscott SJ, González-Prieto R, Chang JG, Vertegeal ACO, Balog J, van der Maarel SM. A proteomics study identifying interactors of the FSHD2 gene product SMCHD1 reveals RUVBL1-dependent DUX4 repression. *Sci Rep*. 2021 Dec 8;11(1):23642. doi: 10.1038/s41598-021-03030-3. PMID: 34880314; PMCID: PMC8654949. (2021).
3. Wang, L. H. & Tawil, R. Facioscapulohumeral Dystrophy. *Curr. Neurol. Neurosci. Rep.* 16, 66 (2016).
4. Deenen, J. C. W. et al. Population-based incidence and prevalence of facioscapulohumeral dystrophy. *Neurology* 83, 1056–9 (2014).
5. Lu-Nguyen N, Malerba A, Antoni Pineda M, Dickson G, Poppelwell L. Improving Molecular and Histopathology in Diaphragm Muscle of the Double Transgenic ACTA1-MCM/FLExDUX4 Mouse Model of FSHD with Systemic Antisense Therapy. *Hum Gene Ther*. 2022 Apr 28. doi: 5.1089/hum.2021.251. Epub ahead of print. PMID: 35078334. (2022).
6. Teeslink S, Vincenten SCC, Voermans NC, Groothuis JT, Doorduyn J, Wijkstra PJ, Horlings CGC, van Engelen BGM, Mul K. Long-term follow-up of respiratory function in facioscapulohumeral muscular dystrophy. *J Neurol*. 2022 Feb 11:1–8. doi: 10.1007/s00415-022-10990-7. Epub ahead of print. PMID: 35147730; PMCID: PMC8831680. (2022).
7. Henke C, Spiesshoefer J, Kabitz HJ, Herkenrath S, Randerath W, Brix T, Görlich D, Young P, Boentert M. Respiratory muscle weakness in facioscapulohumeral muscular dystrophy. *Muscle Nerve*. 2019 Dec;60(6):679-686. doi: 10.1002/mus.26717. Epub 2019 Oct 23. PMID: 31566774. (2019).
8. Fuccillo E, Frezza E, Massa R, Di Girolamo S. Response To Letter To The Editor "Auditory Dysfunction In Facioscapulohumeral Muscular Dystrophy Type 1: Beyond The Inner Ear Involvement" By Gheller et al. *Otol Neurotol*. 2022 Mar 1;43(3):e392-e393. doi: 10.1097/MAO.0000000000003423. PMID: 34772889. (2022).
9. van Dijk GP, van der Kooij E, Behin A, Smeets J, Timmermans J, van der Maarel S, Padberg G, Voermans N, van Engelen B. High prevalence of incomplete right bundle branch block in facioscapulohumeral muscular dystrophy without cardiac symptoms. *Funct Neurol*. 2014 Jul-Sep;29(3):159-65. PMID: 25473735; PMCID: PMC4264782. (2014).
10. Ducharme-Smith A, Nicolau S, Chahal CAA, Ducharme-Smith K, Rehman S, Jaliparthi K, Khan N, Scott CG, St Louis EK, Liewluck T, Somers VK, Lin G, Brady PA, Milone M. Cardiac Involvement in Facioscapulohumeral Muscular Dystrophy (FSHD). *Front Neurol*. 2021 May 24;12:668180. doi: 10.3389/fneur.2021.668180. (2021).
11. Blokhuis AM, Deenen JCW, Voermans NC, van Engelen BGM, Kievit W, Groothuis JT. The socioeconomic burden of facioscapulohumeral muscular dystrophy. *J Neurol*. 2021 May 27. doi: 10.1007/s00415-021-10591-w. (2021).
12. Wallace B, Smith KT, Thomas S, Conway KM, Westfield C, Andrews JG, Weinert RO, Do TQN, Street N; Muscular Dystrophy Surveillance, Tracking, and Research Network (MD STARnet). Characterization of individuals with selected muscular dystrophies from the expanded pilot of the Muscular Dystrophy Surveillance, Tracking and Research Network (MD STARnet) in the United States. *Birth Defects Res*. 2021 Apr 15;113(7):560-569. doi: 10.1002/bdr2.1764. (2020).
13. Mocciaro E, Runfola V, Ghezzi P, Pannese M, Gabellini D. DUX4 Role in Normal Physiology and in FSHD Muscular Dystrophy. *Cells*. 2021 Nov 26;10(12):3322. doi: 10.3390/cells10123322. PMID: 34943834; PMCID: PMC8699294. (2021).
14. Jagannathan S. The evolution of DUX4 gene regulation and its implication for facioscapulohumeral muscular dystrophy. *Biochim Biophys Acta Mol Basis Dis*. 2022 May 1;1868(5):166367. doi: 10.1016/j.bbdis.2022.166367. Epub 2022 Feb 11. PMID: 35158020. (2022).
15. Vuoristo S, Bhagat S, Hydén-Granskog C, Yoshihara M, Gawryski L, Jouhilahti EM, Ranga V, Tamirat M, Huhtala M, Kirjanov I, Nykänen S, Krjutškov K, Damdimopoulos A, Weltner J, Hashimoto K, Recher G, Ezer S, Paluoja P, Paloviita P, Takegami Y, Kanemaru A, Lundin K, Airenne TT, Otonkoski T, Tapanainen JS, Kawaji H, Murakawa Y, Bürglin TR, Varjosalo M, Johnson MS, Tuuri T, Katayama S, Kere J. DUX4 is a multifunctional factor priming human embryonic genome activation. *iScience*. 2022 Mar 22;25(4):104137. doi: 10.1016/j.isci.2022.104137. PMID: 35402882; PMCID: PMC8990217. (2022).
16. Wong CJ, Whiddon JL, Langford AT, Belleville AE, Tapscott SJ. Canine DUXC: Implications for DUX4 retrotransposition and preclinical models of FSHD. *Hum Mol Genet*. 2021 Dec 9;30(24):4352. doi: 10.1093/hmg/ddab352. Epub ahead of print. PMID: 34888646. (2021).
17. Hendrickson, P. G. et al. Conserved roles of mouse DUX and human DUX4 in activating cleavage-stage genes and MERVL/HERVL retrotrans. *Nat. Genet.* 49, 925–934 (2017).
18. Whiddon, J. L., Langford, A. T., Wong, C.-J., Zhong, J. W. & Tapscott, S. J. Conservation and innovation in the DUX4-family gene network. *Nat. Genet.* 49, 935–940 (2017).
19. De Iaco, A. et al. DUX-family transcription factors regulate zygotic genome activation in placental mammals. *Nat. Genet.* 49, 941–945 (2017).
20. Tawil, R. et al. A pilot trial of prednisone in facioscapulohumeral muscular dystrophy. *FSHDY Group. Neurology* 48, 46–9 (1997).

21. Passerieux, E. et al. Effects of vitamin C, vitamin E, zinc gluconate, and selenomethionine supplementation on muscle function and oxidative stress biomarkers in patients with facioscapulohumeral dystrophy: a double-blind randomized controlled clinical trial. *Free Radic. Biol. Med.* 81, 158–69 (2015).
22. Kissel, J. T. et al. Randomized, double-blind, placebo-controlled trial of albuterol in facioscapulohumeral dystrophy. *Neurology* 57, 1434–40 (2001).
23. Elsheikh, B. H. et al. Pilot trial of diltiazem in facioscapulohumeral muscular dystrophy. *Neurology* 68, 1428–9 (2007).
24. Wagner, K. R. et al. A phase 1/II trial of MYO-029 in adult subjects with muscular dystrophy. *Ann. Neurol.* 63, 561–71 (2008).
25. Statland JM, Campbell C, Desai U, Karam C, Diaz-Manera J, Gupta JT, Korngut L, Genge A, Tawil RN, Elman L, Joyce NC, Wagner KR, Manousakis G, Amato AA, Butterfield RJ, Shieh PB, Wicklund M, Gamez J, Bodkin C, Pestronk A, Johnson NE, Mathews KD, Miller B, Leneus A, Fowler M, van de Rijn M, Attie KM. Randomized phase 2 study of ACE-083, a muscle-promoting agent, in facioscapulohumeral muscular dystrophy. *Muscle Nerve.* 2022 Apr 15. doi: 10.1002/mus.27558. Epub ahead of print. PMID: 35428982. (2022).
26. Mellion ML, Ronco L, Berends CL, Pagan L, Brooks S, van Esdonk MJ, van Brummelen EMJ, Oduyungbo A, Thompson LA, Hage M, Badrising UA, Raines S, Tracewell WG, van Engelen B, Cadavid D, Groenewald GJ. Phase 1 clinical trial of losmapimod in facioscapulohumeral dystrophy: Safety, tolerability, pharmacokinetics, and target engagement. *Br J Clin Pharmacol.* 2021 Dec;87(12):4658-4669. doi: 10.1111/bcp.14884. Epub 2021 May 14. PMID: 33931884. (2021).
27. Ghasemi M, Emerson CP Jr, Hayward LJ. Outcome Measures in Facioscapulohumeral Muscular Dystrophy Trials. *Cells.* 2022 Feb 16;11(4):687. PMID: 35203336; PMCID: PMC8870318. (2022).
28. Gros M, Nunes AM, Daoularian D, Pini J, Martinuzzi E, Barbosa S, Ramirez M, Puma A, Villa L, Cavalli M, Grecu N, Garcia J, Siciliano G, Solé G, Juntas-Morales R, Jones PL, Jones T, Glaichenhaus N, Sacconi S. Identification of Serum Interleukin 6 Levels as a Disease Severity Biomarker in Facioscapulohumeral Muscular Dystrophy. *J Neuromuscul Dis.* 2022;9(1):83-93. PMID: 34459413; PMCID: PMC8842759. (2022).
29. Rickard, A. M., Petek, L. M. & Miller, D. G. Endogenous DUX4 expression in FSHD myotubes is sufficient to cause cell death and disrupts RNA splicing and cell migration pathways. *Hum. Mol. Genet.* 24, 5901–14 (2015).
30. Sandri, M. et al. Caspase 3 expression correlates with skeletal muscle apoptosis in Duchenne and facioscapulohumeral muscular dystrophy. A potential target for pharmacological treatment? *J. Neuropathol. Exp. Neurol.* 60, 302–12 (2001).
31. Block, G. J. et al. Wnt/ $\beta$ -catenin signaling suppresses DUX4 expression and prevents apoptosis of FSHD muscle cells. *Hum. Mol. Genet.* 22, 4661–72 (2013).
32. Statland, J. M. et al. Immunohistochemical Characterization of Facioscapulohumeral Muscular Dystrophy Muscle Biopsies. *J. Neuromuscul. Dis.* 2, 291–299 (2015).
33. Kowalow, V. et al. The DUX4 gene at the FSHD1A locus encodes a pro-apoptotic protein. *Neuromuscul. Disord.* 17, 611–23 (2007).
34. Bosnakovski, D. et al. An isogenic myoblast expression screen identifies DUX4-mediated FSHD-associated molecular pathologies. *EMBO J.* 27, 2766–79 (2008).
35. Wallace, L. M. et al. DUX4, a candidate gene for facioscapulohumeral muscular dystrophy, causes p53-dependent myopathy in vivo. *Ann. Neurol.* 69, 540–52 (2011).
36. Geng, L. N. et al. DUX4 activates gemline genes, retroelements, and immune mediators: implications for facioscapulohumeral dystrophy. *Dev. Cell* 22, 38–51 (2012).
37. Yao, Z. et al. DUX4-induced gene expression is the major molecular signature in FSHD skeletal muscle. *Hum. Mol. Genet.* 23, 5342–52 (2014).
38. Homma, S., Beermann, M., Boyce, F. M. & Miller, J. B. Expression of FSHD-related DUX4-FL alters proteostasis and induces TDP-43. *Ann. Clin. Transl. Neurol.* 2, 151–66 (2015).
39. Jagannathan, S. et al. Model systems of DUX4 expression recapitulate the transcriptional profile of FSHD cells. *Hum. Mol. Genet.* 25, 4419–4431 (2016).
40. Jones, T. I. et al. Facioscapulohumeral muscular dystrophy family studies of DUX4 expression: evidence for disease modifiers and a model of pathogenesis. *Hum. Mol. Genet.* 21, 4419–30 (2012).
41. Campbell AE, Shadle SC, Jagannathan S, Lim JW, Resnick R, Tawil R, van der Maarel SM, Tapscott SJ. NuRD and CAF-1-mediated silencing of the D4Z4 array is modulated by DUX4-induced MBD3L proteins. *Elife.* 2018 Mar 13;7. pii: e31023. doi: 10.7554/eLife.31023 (2018).
42. Kolata, G., Reanimated 'Junk' DNA Is Found to Cause Disease. *New York Times, Science.* Published online: August 19, 2010  
<http://www.nytimes.com/2010/08/20/science/20gene.html> (2010).
43. Nurk S, Koren S, Rhie A, Rautiainen M, Bizkadez AV, Mikheenko A, Vollger MR, Altemose N, Uralsky L, Gershman A, Aganezov S, Hoyt SJ, Diekhans M, Logsdon GA, Alongo M, Antonarakis SE, Borchers M, McCoy RC, Dennis MY, Alexandrov LA, Gerton JL, O'Neill RJ, Timp W, Zook JM, Schatz MC, Eichler EE, Miga KH, Phillippy AM. The complete sequence of a human genome. *Science.* 2022 Apr;376(6588):44-53. doi: 10.1126/science.aba6987. Epub 2022 Mar 31. PMID: 35357919 (2022).
44. Jagannathan S1,2,3, Ogata Y4, Gafken PR4, Tapscott SJ3, Bradley RK1. Quantitative proteomics reveals key roles for post-transcriptional gene regulation in the molecular pathology of facioscapulohumeral muscular dystrophy. *Elife.* 2019 Jan 15;8. pii: e41740. doi: 10.7554/eLife.41740 (2019).
45. Bouwman LF, den Hamer B, van den Heuvel A, Tapscott SJ, Rigo F, van der Maarel SM, de Greef JC. Systemic delivery of a DUX4-targeting antisense oligonucleotide to treat facioscapulohumeral muscular dystrophy. *Mol Ther Nucleic Acids.* 2021 Sep 27;26:813-827. doi: 10.1016/j.omtn.2021.09.010. PMID: 34729250; PMCID: PMC8526479. (2021).
46. Soliman HAN, Toso EA, Darwish IF, Ali SM, Kyba M. Antiapoptotic Protein FAIM2 is targeted by miR-3202, and DUX4 via TRIM21, leading to cell death and defective myogenesis. *Cell Death Dis.* 2022 Apr 25;13(4):405. doi: 10.1038/s41419-022-04804-x. PMID: 35468884; PMCID: PMC9038730. (2022).
47. Guo D, Daman K, Chen JJ, Shi MJ, Yan J, Matijasevic Z, Zhou H, Bang AG, Wagner KR, Maehr R, King OD, Hayward LJ, Emerson CP Jr. iMyoblasts for ex vivo and in vivo investigations of human myogenesis and disease modeling. *Elife.* 2022 Jan 25;11:e70341. doi: 10.7554/eLife.70341. PMID: 35076017; PMCID: PMC8789283. (2022).
48. Jagannathan S. The evolution of DUX4 gene regulation and its implication for facioscapulohumeral muscular dystrophy. *Biochim Biophys Acta Mol Basis Dis.* 2022 May 1;1868(5):166367. doi: 10.1016/j.bbdis.2022.166367. Epub 2022 Feb 11. PMID: 35158020; PMCID: PMC9173005. (2022)
49. Mastekia IF, Sathya A, Homma S, Miller BM, Boyce FM, Miller JB. Downstream events initiated by expression of FSHD-associated DUX4: Studies of nucleocytoplasmic transport,  $\gamma$ H2AX accumulation, and Bax/Bak-dependence. *Biol Open.* 2022 Feb 15;11(2):.10.1242/bio.059145. Epub 2022 Feb 22. PMID: 35191484; PMCID: PMC8890089. (2022)
50. Williams K, Yokomori K, Mortazavi A. Heterogeneous Skeletal Muscle Cell and Nucleus Populations Identified by Single-Cell and Single-Nucleus Resolution Transcriptome Assays. *Front Genet.* 2022 May 13;13:835099. doi: 10.3389/fgene.2022.835099. PMID: 35646075; PMCID: PMC9136090. (2022)
51. Hameda CL, Jones PL. FSHD Therapeutic Strategies: What Will It Take to Get to Clinic? *J Pers Med.* 2022 May 25;12(6):865. PMID: 35743650; PMCID: PMC9225474. (2022)
52. Lu-Nguyen N, Dickson G, Malerba A, Poppellwell L. Long-Term Systemic Treatment of a Mouse Model Displaying Chronic FSHD-like Pathology with Antisense Therapeutics That Inhibit DUX4 Expression. *Biomedicines.* 2022 Jul 7;10(7):1623. doi: 10.3390/biomedicines10071623. PMID: 35884928; PMCID: PMC9313434. (2022)
53. Mariot V, Dumonceaux J. Gene Editing to Tackle Facioscapulohumeral Muscular Dystrophy. *Front Genom Ed.* 2022 Jul 15;4:937879. doi: 10.3389/fgene.2022.937879. PMID: 35910413; PMCID: PMC9334676. (2022)
54. Azzag K, Bosnakovski D, Tungtur S, Salama P, Kyba M, Perlinger RCR. Transplantation of PSC-derived myogenic progenitors counteracts disease phenotypes in FSHD mice. *NPJ Regen Med.* 2022 Sep 27;7(1):43. doi: 10.1038/s41536-022-00249-0. PMID: 36056021; PMCID: PMC9440030. (2022)
55. Hiramuki Y, Kure Y, Saito Y, Ogawa M, Ishikawa K, Mori-Yoshimura M, Oya Y, Takahashi Y, Kim DS, Arai N, Mori C, Matsumura T, Hamano T, Nakamura K, Ikezoe K, Hayashi S, Goto Y, Noguchi S, Nishino I. Simultaneous measurement of the size and methylation of chromosome 4qA-D4Z4 repeats in facioscapulohumeral muscular dystrophy by long-read sequencing. *J Transl Med.* 2022 Nov 8;20(1):517. doi: 10.1186/s12967-022-03743-7. PMID: 36348371; PMCID: PMC9644496. (2022)
56. Lim KRQ, Yokota T. Knocking Down DUX4 in Immortalized Facioscapulohumeral Muscular Dystrophy Patient-Derived Muscle Cells. *Methods Mol Biol.* 2023;2587:197-208. doi: 10.1007/978-1-0716-2772-3\_12. PMID: 36401032. (2023)
57. Heskamp L, Ogier A, Bendahan D, Heerschap A. Whole-muscle fat analysis identifies distal muscle end as disease initiation site in facioscapulohumeral muscular dystrophy. *Commun Med (Lond).* 2022 Dec 1;2(1):155. doi: 10.1038/s43856-022-00217-1. Erratum in: *Commun Med (Lond).* 2023 Jan 12;3(1):6. PMID: 36450865; PMCID: PMC9712512. (2023)
58. Bosnakovski D, Toso EA, Ener ET, Gearhart MD, Yin L, Lüttmann FF, Magli A, Shi K, Kim J, Aihara H, Kyba M. Antagonism among DUX family members evolved from an ancestral toxic single homeodomain protein. *bioRxiv [Preprint].* 2023 Jan 22:2023.01.21.524976. doi: 10.1101/2023.01.21.524976. PMID: 36711898; PMCID: PMC9882399. (2023)
59. Nip Y, Bennett SR, Smith AA, Jones TL, Jones PL, Tapscott SJ. Human DUX4 and porcine DUXC activate similar early embryonic programs in pig muscle cells: implications for preclinical models of FSHD. *Hum Mol Genet.* 2023 Feb 2;ddad021. doi: 10.1093/hmg/ddad021. Epub ahead of print. PMID: 36728804. (2023)
60. Wong CJ, Friedman SD, Snider L, Bennett SR, Jones TL, Jones PL, Shaw DWW, Blemker SS, Riem L, DuCharme O, Lemmers RJFL, van der Maarel SRM, Wang LH, Tawil R, Statland JM, Tapscott SJ. Validation of the association between MRI and gene signatures in facioscapulohumeral dystrophy muscle: implications for clinical trial design. *bioRxiv [Preprint].* 2023 Feb 20:2023.02.20.529303. doi: 10.1101/2023.02.20.529303. PMID: 36865168; PMCID: PMC9980042. (2023)
61. Claus C, Slavin M, Anseau E, Lancelot C, Bah K, Lassche S, Fiévet M, Greco A, Tomaiuolo S, Tassin A, Dudome V, Kusters B, Declèves AE, Laoudj-Chenivisse D, van Engelen BGM, Nonclercq D, Belayev A, Kalisman N, Coppée F. The double homeodomain protein DUX4c is associated with regenerating muscle fibers and RNA-binding proteins. *Skelet Muscle.* 2023 Mar 7;13(1):5. doi: 10.1186/s13395-022-00310-y. PMID: 36882853; PMCID: PMC9990282. (2023)
62. Padberg GW, van Engelen BGM, Voermans NC. Facioscapulohumeral Disease as a myodevelopmental disease: Applying Ockham's razor to its various features. *J Neuromuscul Dis.* 2023;10(3):411-425. doi: 10.3233/JND-221624. PMID: 36872787. (2023)
63. Mocciano E, Giambruno R, Micheloni S, Cemilogar FM, Andolfo A, Consonni C, Pannese M, Ferri G, Runfola V, Schotta G, Gabellini D. WDR5 is required for DUX4 expression and its pathological effects in FSHD muscular dystrophy. *Nucleic Acids Res.* 2023 Apr 6;gkad230. doi: 10.1093/nar/gkad230. Epub ahead of print. PMID: 37021550. (2023)
64. Spens AE, Sutliff NA, Bennett SR, Campbell AE, Tapscott SJ. Human DUX4 and mouse Dux interact with STAT1 and broadly inhibit interferon-stimulated gene induction. *Elife.* 2023 Apr 24;12:e82057. doi: 10.7554/eLife.82057. PMID: 37092726. (2023)
65. Knox RN, Eidahl JO, Wallace L, Choudury S, Rashnoejad A, Daman K, Guggenbiller M, Saad N, Hoover ME, Zhang L, Branson OE, Emerson CP Jr, Freitas MA, Harper SQ. Posttranslational modifications of the DUX4 protein impact toxic function. *Ann Neurol.* 2023 Apr 26. doi: 10.1002/ana.26668. Epub ahead of print. PMID: 37186119. (2023)
66. Ganassi M, Zammit PS. Involvement of muscle satellite cell dysfunction in neuromuscular disorders: Expanding the portfolio of satellite cellopathies. *Eur J Transl Myol.* 2022 Mar 18;32(1):10064. doi: 10.4081/ejtm.2022.10064. PMID: 35302338; PMCID: PMC8992676. (2022)
67. Cowley MV, Pruller J, Ganassi M, Zammit PS, Banerji CRS. An in silico FSHD muscle fibre for modelling DUX4 dynamics and predicting the impact of therapy. *Elife.* 2023 May 15;12:e88345. doi: 10.7554/eLife.88345. Epub ahead of print. PMID: 37184373. (2023)